Abstract—Antifungal activities of ether and methanolic extracts of volatiles oils of Nigella Sativa seeds were tested against pathogenic bacteria and fungi strains. The volatile oil were found to have significant antifungal and antibacterial activities compared to tetracycline, cefuroxime and ciprofloxacin positive controls. The ether and methanolic extracts were compared to each other for antifungal and antibacterial activities and ether extracts showed stronger activity than methanolic one.

Keywords—Antifungal, antibacterial, essential oils, extraction, Nigella Sativa.

I. INTRODUCTION

A large number of medicinal plants have therapeutic potentials. Seeds of Nigella sativa L. (Ranunculaceae), known commonly as “black cumin” have been employed for thousands years as a spice and food preservative. The oil and seed constituents have shown potential medicinal properties in traditional medicine. Recently, many biological activities of Nigella sativa L. seeds have been reported, including: antioxidant, anti-inflammatory, anticancer and antimicrobial and antifungal ones. [2]

Several pharmacological effects have been attributed to active principles of Nigella sativa L. which includes thymoquinone, thymohydroquinone, dithymoquinone, thymol, carvacrol, nigellicine, nigellimine-x-oxide, nigellidine and alpha-hedrin (Aljabre et al. 2005). [3],[7],[11] Nigella sativa L. seed extract inhibits fungal strains. In our study we have tested the antifungal and antibacterial properties of Nigella Sativa.

II. EXPERIMENTS

A. Extraction of the Essential Oils

25 g seeds were crushed and extracted with petroleum ether for 4 h in a Soxhlet apparatus. After extraction, the solvents were removed by rotary vacuum and dried in a vacuum oven at 30°C for. The same method was repeated by using methanol as extract agent.

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B. Materials and Methods


C. Antimicrobial and Antifungal Activity of Essential Oil

The essential oil of samples M1- Ether extract and M2- methanol extract were tested for antibacterial activity by the disc diffusion method using 100µL of suspension of the tested microorganisms, containing 2.0 x 106 colony forming units (cfu mL–1) for bacteria and 2.0 x 105 spore mL–1 for fungal strains. [11]

Fig.1 Essential Oils Inhibition Zone And Positive Controls vs /C. Albicans

Mueller–Hinton agar and dextrose agar were distributed to sterilized Petri dishes with a diameter of 9 cm.

The filter paper discs (6 mm in diameter) were individually impregnated with 10µL and 30µL of the essential oils dissolved in dimethylsulfoxide (DMSO). (Fig 1, Fig 2)

The Petri dishes were kept at 4°C for 2 h. The plates inoculated with bacteria incubated at 37°C for 24 h and at 30 °C for 48 h for the yeasts. The diameters of the inhibition zones were measured in millimetres. Negative controls were set up with equivalent quantities of DMSO. Studies were performed in triplicate. In addition, positive controls antibiotic discs such as Cefuroxime, Ciprofloxacin, Tetracycline and Nystatin were used for comparison.

II. RESULTS AND DISCUSSION

The results of disc diffusion assay are demonstrated on tab I.
III. DISCUSSION

*Nigella Sativa* essential oils are more sensible against gram positive bacteria. (Graph 1) then those gram - negative ones. (Graph 2,3)

*Nigella Sativa* essential oil have stronger antibacterial properties compare to Cefuroxime, Tetracycline, (Graph 1,2) and about the same strength compare to ( Graph 3) Ciprofloxacine and antifungal properties compare to Clomatrizol. (Graph 4)

*Nigella Sativa* essential oils with high concentration of (30 ug) carvacrol and thymol are more sensible against bacterias then those with lower concentrations (10 ug) maybe because they are responsible of the antibacterial activity. (Graph 1,2,3,4 and fig 1,2)

*Nigella Sativa* essential oils Methanol extracts have more antibacterial and antifungal properties than etheric ones.

IV. CONCLUSION

It may be concluded from this study that N. sativa seed extract has antimicrobial activity against Staphylococcus Aerus, Proteus Vulgaris, Escherichia Coli, Candida Albicans. It is expected that using natural products as therapeutic agents will probably not elicit resistance in microorganisms. It is essential that research should continueto isolate and purify the active components of this natural herb and use in experimental animals.

REFERENCES


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**Table I** BACTERIA AND FUNGI MEAN OF INHIBITION ZONE OF ETHER AND METHANOLIC EXTRACTS

<table>
<thead>
<tr>
<th>Bacteria and Fungi</th>
<th>Ether extracts mean inhibition zone (mm)</th>
<th>Methanolic extracts mean inhibition zone. (mm)</th>
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<tbody>
<tr>
<td>Staphylococcus Aureus ATCC 29737 Lot 58312397</td>
<td>37±0.1</td>
<td>38±0.3</td>
</tr>
<tr>
<td>Proteus Vulgaris ATCC 1978 Lot 0876523C</td>
<td>31±0.8</td>
<td>29±0.6</td>
</tr>
<tr>
<td>Escherichia Coli, ATCC 8456 LOT 6543109</td>
<td>21±0.7</td>
<td>23±0.4</td>
</tr>
<tr>
<td>Candida Albicans, ATCC 2091 Lot 7051869</td>
<td>21±0.5</td>
<td>24±0.8</td>
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