Mathematical modeling of Bi-Substrate Enzymatic Reactions with Ping-Pong Mechanism in the Presence of Competitive Inhibitors

Rafayel A. Azizyan, Aram E. Gevorgyan, Valeri B. Arakelyan, and Emil S. Gevorgyan

Abstract—The mathematical modeling of different biological processes is usually used to predict or assess behavior of systems in which these processes take place. This study deals with mathematical and computer modeling of bi-substrate enzymatic reactions with ping-pong mechanism, which play an important role in different biochemical pathways. Besides that, three models of competitive inhibition were designed using different software packages. The main objective of this study is to represent the results from in silico investigation of bi-substrate enzymatic reactions with ordered ping-pong mechanism in the presence of competitive inhibitors, as well as to describe in details the inhibition effects. The simulation of the models with certain kinetic parameters allowed investigating the behavior of reactions as well as determined some interesting aspects concerning influence of different cases of competitive inhibition. Simultaneous presence of two inhibitors, competitive to the $S_1$ and $S_2$ substrates have been studied. Moreover, we have found the pattern of simultaneous influence of both inhibitors.

Keywords—Mathematical modeling, bi-substrate enzymatic reactions, ping-pong mechanism, competitive inhibition.

I. INTRODUCTION

Enzymatic reactions with participation of two substrates are called bi-substrate enzymatic reactions and are widely spread in different metabolic pathways from simple organisms to highly developed ones [1]-[3], [12]. There are several well-known mechanisms of bi-substrate enzymatic reactions, namely sequential mechanism, ping-pong mechanism and iso-mechanism [3], [4], [6].

In the case of ping-pong mechanism product is already released before all substrates are bound. The ping-pong mechanism can be categorized into two groups, namely random ping-pong and ordered ping-pong mechanisms. In contrast to the random mechanism, during the ordered mechanisms substrates bind to the enzyme in a defined order.

According to the Cleland’s schematic representation of enzymatic reactions, different states of the enzyme can be represented by a horizontal line and the substrates and products by vertical arrows [5]. Thus, the scheme for bi-substrate ping-pong enzymatic reaction is as follows:

$$
E \xrightarrow{k_1} [ES_1] \xrightarrow{k_2} F \xrightarrow{k_3} [FS_2] \xrightarrow{k_4} E
$$

where $k_1; k_2; k_3; k_4$ and $k_5; k_5$ are rate constants of forward and reverse reactions, respectively; $E$ is concentration of free enzyme; $S_1$ and $S_2$ are concentrations of the first and the second substrates, respectively; $[ES_1]$ represents binary complex $(E-S_1)$; $[FS_2]$ is for binary complex $(F-S_2)$; $P_1$ and $P_2$ are the first and the second products of enzymatic reaction, respectively.

The following system of differential equations describes the bi-substrate ping-pong enzymatic reactions [7], [11]:

$$
dE/dt = k_{-1}[ES_1] + k_4[FS_2] - k_1[E][S_1] \\
d[S_1]/dt = k_{-1}[ES_1] - k_1[E][S_1] \\
d[ES_1]/dt = k_1[E][S_1] - k_{-1}[ES_1] - k_2[ES_1] \\
dP_1/dt = k_2[ES_2] \\
dF/dt = k_3[ES_1] + k_{-3}[FS_2] - k_3[F][S_2] \\
d[S_2]/dt = k_{-3}[FS_2] - k_3[F][S_2] \\
d[FS_2]/dt = k_3[F][S_2] - k_{-3}[FS_2] - k_4[FS_2] \\
dP_2/dt = k_4[FS_2]
$$

An enzyme inhibitor is a compound that binds to an enzyme and interferes with its activity, consequently by slowing down, or in some cases, stopping the catalysis [12]. Cells contain many natural enzyme inhibitors that play important roles in regulating metabolism. Artificial inhibitors are used experimentally to investigate enzyme mechanisms and decipher metabolic pathways. Some drugs, and many poisons, are enzyme inhibitors too [11].

Some inhibitors bind covalently to enzymes causing irreversible inhibition but most biologically relevant inhibition is reversible. Reversible inhibitors are bound to enzymes by the same weak, non-covalent forces that bind substrates and products. Three common types of reversible enzyme inhibition are known in literature: competitive, noncompetitive and uncompetitive inhibition. Here we discuss only competitive inhibition.

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Competitive inhibition occurs when the substrate and an inhibitor resembling the substrate are both added to the enzyme. Once a competitive inhibitor is bound to an enzyme molecule, a substrate molecule cannot bind to that enzyme molecule. Conversely, the binding of substrate to an enzyme molecule prevents the binding of an inhibitor. In other words, S and I compete for binding to the enzyme molecule.

For bi-substrate enzymatic reactions with ping-pong mechanism the following three cases of inhibition are studied:

- Competitive inhibition to the first $S_1$ substrate – designate PPM1 (ping-pong model-1).
- Competitive inhibition to the second $S_2$ substrate – designate PPM2 (ping-pong model-2).
- Simultaneous presence of both inhibitors – designate PPM3 (ping-pong model-3).

Schematically, these three cases of inhibition can be represented as follows:

For bi-substrate enzymatic reactions with ping-pong mechanism the following three cases of inhibition are studied:

II. METHODS

Two different modeling software packages are used to design the three models for each above mentioned cases for ordered ping-pong mechanism. Modeling has been carried out using “STELLA” dynamic modeling package and “Mathematica” software based on the above-presented order differential equations (ODEs) [8], [9]. In “STELLA” the computing was done by Euler’s method of integration while in “Mathematica 7” the Runge-Kutta’s method of integration was used.

Since the duration of real biological reactions does not correspond to the model simulation time, the description of kinetic behavior of models has done based on conditional time units.

The following same kinetic parameters are used in all models:

$$E_0=10 \, \mu\text{mol} \quad k_1=2\times10^{-3} \, (\text{sec}\times\mu\text{mol})^{-1} \quad k_{-1}=1\times10^{-3} \, (\text{sec})^{-1}$$

$$S_1=300 \, \mu\text{mol} \quad k_2=5\times10^{-3} \, (\text{sec})^{-1} \quad k_i=k_j=10^{-3} \, (\text{sec}\times\mu\text{mol})^{-1}$$

$$I=30 \, \mu\text{mol} \quad k_4=5\times10^{-3} \, (\text{sec})^{-1} \quad k_{-i}=k_{-j}=7\times10^{-4} \, (\text{sec})^{-1}$$

(4) and (8) correspond to $P_1$ and $P_2$ products generation respectively, while substrates consumption determined by (2) and (6).

III. RESULTS AND DISCUSSION

The simulation of the above mentioned models by both software packages led to similar outcomes. The results of the simulations were discussed below in terms of separate parameters.

![Fig. 1 Concentration changes of the first $P_1$ (part A) and the second $P_2$ (part B) products during enzyme kinetics. Curve 1 corresponds to the concentration changes of $P_1$ and $P_2$ from baseline model, without any inhibitors, curve 2 – model with competitive inhibitor to $S_1$ and curve 3 – competitive to $S_2$ substrate](image)

It is natural, that dynamics of the change in product concentration shows significant decrease in the rate of product generation in the presence of any inhibitors compared to the baseline case, when no inhibitor is presented (Fig. 1).

As one can notice on Fig. 1, the influence of the competitive inhibitor to the first substrate (curve-2) is a little bit more effective in terms of decreasing the rate of product generation than one to the second substrate (curve-3). This is a result of the fact that the inhibitor to the first substrate binds to the enzyme a little bit earlier than the competitive inhibitor to the second substrate. In other words, the rate of formation $[ES_1]$ and $[FS_2]$ catalytic complexes decrease and the overall enzymatic reaction rate goes down [10].
The more interesting and essential task is the presence of both inhibitors in virtual solution. The question is, how does this process occur? What is the law of inhibition effect in the presence of both inhibitors: is it additive, whether it is less than in the case of separate presence of inhibitors, or maybe it synergistic?

To find answers to the above questions, we have constructed another model (using the same initial kinetic parameters), corresponding to the simultaneous presence of two different competitive inhibitors, for simulation in both software packages. As simulations show, and as it was expected, the inhibition effect of both inhibitors in simulta neous presence is more than inhibition effect of same inhibitors taking separately. This inhibitory effect illustrated in Fig. 2.

From Fig. 2, clearly seen, that curve corresponding to enzyme kinetics in the presence of two different competitive inhibitors (curve-4), lies under the other curves, which means that inhibition effect is the most significant.

For numerical evaluation and comparison of inhibition effects in all studied models, we suggested to represent all derived data corresponding to the time conditional time units (CTUs), when release of products tends to be maximum possible one, in baseline model, where no inhibitor presents. Particularly, that time points correspond to the 1851th, 4035th and 6724th conditional time units, respectively (Table I).

<table>
<thead>
<tr>
<th>Models</th>
<th>Values of products concentration (µmol) in different models at the end of baseline model kinetics</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>P1 (at 14280th CTU) Consumption (%)</td>
</tr>
<tr>
<td>Baseline model</td>
<td>299.99</td>
</tr>
<tr>
<td>PPM1</td>
<td>256.56</td>
</tr>
<tr>
<td>PPM2</td>
<td>274.64</td>
</tr>
<tr>
<td>PPM3</td>
<td>227.60</td>
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</tbody>
</table>

IV. CONCLUSIONS

The following conclusions can be drawn based on the results of simulations:

In terms of product generation of the bi-substrate enzymatic reaction the competitive inhibitor to the first substrate acts more effectively than the competitive inhibitor to the second substrate.

Simultaneous presence of the both competitive inhibitors influence on the dynamics of product generation with significantly different manner. Moreover, that manner is a little more than additive one.

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REFERENCES